1	Running title: ESOPHAGEAL TONSIL OF THE CHICKEN
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3	Title: ESOPHAGEAL TONSIL; A NOVEL GUT-ASSOCIATED LYMPHOID
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26 ABSTRACT

The esophageal tonsil of the chicken is a novel, significant element of the GALT. Its stable location and histological organization fulfills the meaning of the term "tonsil". The six-eight isolated tonsillar units locate at the border of the esophagus and the proventriculus. The number of tonsillar units is identical with that of the esophageal folds. Each tonsillar unit consists of a crypt lined by lymphoepithelium and surrounded by dense lymphoid substance. The lymphoid substance is organized into T and B dependent regions, like peripheral lymphoid organs. The excretory ducts of the mucosal glands of the esophagus are frequently involved into the formation of the lymphoepithelium. The esophageal tonsil anatomically locates before the stomach, unlike the other parts of the GALT, therefore it is continuously exposed to undigested environmental antigens, allergens, food and infectious agents. The endeavor to develop oral vaccination further underline its functional significance.

Keywords: GALT, esophageal tonsil, chicken, lymphoepithelial tissue.

INTRODUCTION

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The avian gut-associated lymphoid tissue (GALT) has been extensively studied (Oláh and Glick, 1978; Befus et al. 1980; Glick and Oláh, 1981; Oláh et al.1984; Dolfi et al. 1988; Vervelde and Jeurissen 1993; Cortes et al. 1995; Lillehoj et al. 1996; Nagy et al. 2001, Yasuda et al. 2002), and reviewed (Payne, 1971; Glick 1988, Jeurissen et al. 1994). Along the entire digestive tract small lymphoid accumulations, primary follicles occur besides the classical lymphoid organs, like the bursa of Fabricius (Bockman and Cooper 1973, Glick 1988, Dasso et al. 2000), caecal tonsils (Oláh and Glick 1979), Peyer's patches (Befus et al. 1980), Meckel's diverticulum (Oláh et al. 1984) and diffusely infiltrated areas of the cloaca (Odend'hal and Breazile, 1980; Gomez del Moral et al. 1998). There is very sketchy histological and no immunological information on the lymphoid substance locating at the border of the esophagus and proventriculus. In the available histological textbook of fowls (Hodges, 1974) only one sentence has been devoted to this lymphoid accumulation; "The junction between the esophagus and the proventriculus is frequently heavily infiltrated with lymphatic cells and there are often many large lymphoid foci lying in the tunica". This is the only reference that is related with the existence of lymphoid tissue at the junction of the esophagus and proventriculus.

The significance to deal with this remarkable lymphoid tissue is double; 1) theoretical: it may be participated in the B cell development and (Ratcliffe 2002), 2) practical: this is the only substantial lymphoid accumulation which locates in the digestive tract before the stomach. Because of its unique location it is constantly exposed by undigested environmental antigens.

Therefore, to be familiar its morphological structure and immunological function could be crucial to understand tolerance to undigested antigens, food allergy and intestinal infection, like infectious bursal disease virus. So, in this short paper we introduce the basic anatomical and histological structure of the lymphatic accumulation at the border of esophagus and proventriculus, what we called esophageal tonsil.

MATERIALS AND METHODS

Animals

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Fertilized White Leghorn SPF chicken eggs were obtained from CEVA-77 Phylaxia Hungary and incubated at 37,7°C in humidified incubator. 78 Histological examinations were made from 4, 6, 8, 12 week old chickens. The 79 design and condition of the animal experiments were approved by the Animal 80 81 Ethical Committee of Semmelweis University, Budapest, Hungary (TUKEB/2000). The animals were sacrificed by cervical distortion. 82

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Antibodies

To identify hemopoietic cells and B lymphocytes anti-chick CD45 monoclonal antibody (mAb, clone: HIS-C7; Jeurissen et al. 1988) and anti-chick Bu1b mAb (clone: 11G2) were used. Anti-CD45 and Bu1b mAb(s) were generous gift from Dr. Haymo Kurz, Albert-Ludwigs University, Freiburg, Germany and Dr. Olli Vainio, Turku University, Finland), respectively. Anti-CD3 mAb (clone: CT3) were obtained from Dr. Chen-Lo Chen, University of Alabama at Birmingham, Birmingham, AL. Epithelial cells were visualized by using a pan-cytokeratin mAb (clone: Lu-5)¹.

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Histological procedures

For cryostat sections the border of oesophagus and proventriculus was cut out and snap frozen in liquid nitrogen. The ten-micrometer frozen sections were collected on poly-L-lisine coated slides (Sigma)², fixed in cold acetone and air dried. For histological examination the tissue samples were fixed in buffered 4% glutaraldehyde (Merck)³ solution for overnight and postfixed in

1% osmium tetroxide (Polyscience)⁴ solution for two hours. After rehydration in graded ethanol, the tissue samples were embedded in Polybed/Araldite 6500 mixture (Polyscience)⁴. The one-micrometer thick semithin sections were stained with toluidine blue. For hematoxilin-eosin staining the specimens were fixed in buffered formalin and embedded in paraffin.

Immunocytochemistry

Immunocytochemistry was made on cryostat sections according to standard techniques. Briefly, after rehydration in PBS, the acetone fixed sections were incubated with primary antibodies for 45 minutes, followed by biotinylated horse anti-mouse IgG (Vector)⁵ and avidin-biotinylated peroxidase complex (ABC, Vectastain Elite ABC kit, Vector)⁵ at room temperature. Endogenous peroxidase activity was quenched before ABC incubation by treatment of the sections with 3% hydrogen peroxid (Sigma)² for 10 minutes. The binding sites of the primary antibodies were visualized by 4-chloro-1-naphthol (Sigma)².

Footnotes:

- ¹BMA, BIOMEDICALS AG, Rheinstrasse 28-32; CH-4302 Augst, Switzerland,
- ²Sigma- Aldrich Kft. Nagydiófa str. 7, H-1072 Budapest, Hungary.
- ³Merck Kft., Talpas str. 3, H-1116 Budapest, Hungary.
- ⁴Polysciences, Inc. 400 Valley Road, Warrington, PA 18976.
- ⁵Vector Laboratories, Inc. 30 Ingold Road, Burlingame, CA 94010

125 RESULTS

The esophagus forms six-eight longitudinal folds, which create a stellate-shaped appearance on cross-section. The lymphoid tissue is localized at the distal ends of the folds, before the esophagus turns to proventriculus (Fig. 1a). This area is about three-eight millimeter long. At this region the tunica propria and the lymphoid tissue are covered by stratified squamous epithelium but the proventricular glands are already emerged. Cylindrical, proventricular epithelium occurs only above the most distal part of the lymphoid tissue (Fig.1b). The lymphoid tissue is associated with the bottom of the folds, where it forms isolated units. Although the lymphoid units appear in circumferencial in the wall of the esophagus, it does not form a continuous ring. The number of the units is identical with that of the longitudinal folds of the esophagus (Fig.1a). This finding suggests, that the esophageal folds are not randomly formed during swallowing, but they are anatomically determined i.e. they are stable structures of the esophagus.

The real tonsillar lacuna or crypt starts at the bottom of the folds and reaches the muscular layer of the tunica mucosa. The tonsillar unit consists of a crypt and the surrounding lymphoid tissue (Fig.1c). Unlike the Peyer's patches, where the lymphoid tissue occupies the tunica propria and submucosal layers of the intestine, these tonsillar units are restricted to the tunica propria (Fig. 1b). In the borderline of the esophagus and proventriculus the proventricular glands gradually fill up the submucosa of the esophagus, while towards the proventriculus the tunica muscularis mucosae breaks up. It is apt to notice, that the proventricular glands reveal scattered primary follicles (Matsumoto and Hashimoto, 2000), which might be involved into the

pathogenesis of a new variant of the infectious bursal disease virus resulting in proventriculitis (Dormitorio et al. 2001; Giambrone, 2002). These primary follicles do not show anatomical relationship with the esophageal tonsil.

Blood-borne lymphoid tissue surrounds the crypts (Fig.1d.) and infiltrates the stratified squamous epithelium (Fig.2a). The infiltration of the surface epithelium turns it to lymphoepithelium (Fig.2b). The heavily stained B lymphocytes are localized in the germinal centers, but many weakly stained Bu-1b positive cells also occur in the T dependent interfollicular areas (Fig.2c). In addition to the T cell regions significant number of T cells also occur in the germinal centers (Fig.2d).

Mucosal glands of the esophagus are frequently associated with the lymphoid substance. The secretory acinus is generally free of mobile cells, but the excretory ducts are involved in the formation of lymphoepithelium. Interesting, that the wall of the duct is covered partly by cylindrical and partly by stratified squamous epithelium, and only the latter part of the duct is infiltrated with lymphoid cells. (Fig.2e). Occasionally the lumen of the duct is dilated and filled with detritus.

DISCUSSION

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The gut-associated lymphoid tissue of the chicken consists of solitary and aggregated lymphoid nodules. The latter appears as lymphoid organs. Now, we add to the GALT a novel organ, which has not yet got enough attention. Possibly, it has been thought, that this lymphoid tissue accumulation represents only small, variable, insignificant lymphoid substance. However, the location of this lymphoid substance is anatomically strictly determined, therefore it is a stable, consistent lymphoid structure, like the caecal tonsil. We named this lymphoid substance as esophageal tonsil.

Generally, the correct term of the tonsil covers a complex organ, which consists of an epithelium lined crypt surrounded by dense lymphoid substance. The epithelium is infiltrated with mobile lymphoid cells creating lymphoepithelial tissue. The lymphoid substance shows a well-organized structure, namely in the interfollicular space mainly T cell are accumulated, while the B cells form germinal centers. The described structural unit is identical with the simplest tonsillar structure, occurring in the human lingual and rabbit palatine tonsils. So, the term "tonsil" to characterize this lymphoid accumulation perfectly fulfills the histological postulate of the tonsil.

Each isolated tonsillar unit is associated to the bottom of two neighboring folds, which suggests, that each unit has separate blood- and lymphatic circulation and form a functional and/or pathological unit.

The esophageal tonsil locates before the stomach, unlike the major parts of the GALT, so it is continuously exposed by large amount of undigested environmental antigens, food and allergens. Due to its location, its function could be delicate comparing with other parts of the GALT, which can be found in the alimentary tract distal to the stomach. This unique anatomical location may be taken into consideration for developing oral vaccination.

The mucosal glands of the lower esophagus frequently associated with the lymphoid substance. The excretory ducts of the glands are partly lined by stratified squamous epithelium, which is transformed to lymphoepithelium. The significance of this association is unknown, but we speculate, that the lymphoepithelial tissue of the tonsil functions as "gate" for environmental antigens, through which antigens and/or allergens continuously stimulate the immune system. The secretion washes away the excess of antigens and allergens. Possibly, the insufficient cleanup function of the gland results in obliteration of the excretory duct, and consequently formation of "salivary bodies".

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208	APPENDIX
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LEGEND OF FIGURES 299 300 Fig 1a. 301 302 HE-stained cross-section of the esophagus shows seven longitudinal folds. At the bottom of the esophagus seven tonsillar units are present in the lamina 303 propria. Mucosal glands of the esophagus (arrow). Mag: x13 304 305 Fig 1b. HE-stained longitudinal section through a tonsillar unit, which is covered by 306 stratified squamous epithelium. A mucosal gland (MG) is embedded into the 307 308 lymphatic substance. The arrow marks the border of the stratified squamous 309 and cylindrical epithelium. Mag: x32 Fig 1c. 310 311 HE-stained tonsillar unit. The lymphoid substance is organized to germinal centers (GC) and interfollicular areas. Mag: x30 312 313 Fig 1d. Anti CD-45 mAb (hemopoietic cell marker) clearly outlines the tonsillar units at 314 315 the bottom of the esophageal folds. A primary follicle is also present in one of 316 the proventricular gland (arrow). Mag: x8 317 Fig 2a. Anti-cytokeratin stained cryostat section shows the stratified squamous 318 319 epithelium of the e esophagus and the associated mucosal glands (arrow). 320 The epithelial cells of the proventricular glands (PG) also express keratin intermediate filaments. Mag: x8 321

- 322 Fig 2b.
- One micron thick section stained with toluidin blue. Surface epithelium is
- 324 heavily infiltrated by lymphoid cells, that transform the stratified squamous
- epithelium to lymphoepithelium. Mag: x40
- 326 Fig 2c.
- 327 Bu-1b mAb identifies a heavily- and a lightly stained cell populations in the
- germinal centers and in interfollicular areas, respectively. Mag: x8
- 329 Fig 2d.
- 330 Anti CD3 (T cell) mAb heavily stains the interfollicular areas and reveals
- significant number of T cells inside the germinal centers (GC). The surface
- epithelium also shows CD3 positive cells. Mag: x30
- 333 Fig 2e.
- HE-stained section shows the duct of a mucosal gland. One part of the duct is
- covered by stratified squamous epithelium which is infiltrated by lymphoid
- cells, while the other part of the duct is lined by cylindrical epithelium. LE,
- 337 Lymphoepithelium. Mag: x144